



## A quantitative structure–retention relationship for the prediction of retention indices of the essential oils of *Ammoides atlantica*

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**Abstract:** A simple, descriptive and interpretable model, based on a quantitative structure–retention relationship (QSRR), was developed using the genetic algorithm–multiple linear regression (GA-MLR) approach for the prediction of the retention indices (*RI*) of essential oil components. By molecular modeling, three significant descriptors related to the *RI* values of the essential oils were identified. A data set was selected consisting of the retention indices for 32 essential oil molecules with a range of more than 931 compounds. Then, a suitable set of the molecular descriptors was calculated and the important descriptors were selected with the aid of the genetic algorithm and multiple regression method. A model with a low prediction error and a good correlation coefficient was obtained. This model was used for the prediction of the *RI* values of some essential oil components which were not used in the modeling procedure.

**Keywords:** chemometrics; QSRR; genetic algorithms; multiple linear regression; retention indices; essential oils.

### INTRODUCTION

Essential oils are valuable natural products used as raw materials in many fields, including folk medicine, perfumes, cosmetics, aromatherapy, phytotherapy, spices and nutrition. They are mixtures of more than 200 compounds that can be grouped basically into two fractions, a volatile fraction, which constitutes 90–95 % of the whole oil and contains monoterpenes and sesquiterpene hydrocarbons and their oxygenated derivatives, together with aliphatic aldehydes, al-

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cohols and esters, and a nonvolatile residue, that constitutes 5–10% of the whole oil and contains hydrocarbons, fatty acids, sterols, carotenoids, waxes, coumarins, psoralens and flavonoids.<sup>1</sup>

*Ammoides atlantica* is of the *Apiaceae* family. The plant is used in infusions for the treatment of headache, fever and diarrhea. It is also used in compresses, alone or soaked in alcohol or vinegar and mixed with henna, to treat children affected by mental debility as reported by Laouer *et al.*<sup>2</sup>

GC and GC–MS are the main methods for the identification of these plant oils. Seeking a quantitative relationship between the molecular structure and the gas chromatographic retention indices has been a basic task in chemistry. Correlations between the GC retention indices and the molecular structures can provide more profound insights into the interactions between the eluents and the stationary phases from a theoretical viewpoint. In addition, they can provide very important information about the effect of the chemical structures on the retention behavior and the possible mechanism of absorption and elution.

Prediction of physico–chemical properties of materials based on their molecular structure has been one of the wishes of scientists and engineers for a long time. One of the best methods which has been applied for this purpose is quantitative structure–property relationships (QSPR). QSPR analysis is now a well-established and highly respected technique to correlate diverse simple and complex physico–chemical properties of a compound with its molecular structure, through a variety of descriptors. The basic strategy of QSPR analysis is to find optimum quantitative relationships, which can then be used for the prediction of the properties from molecular structures. Once a reliable relation has been obtained, it is possible to use it to predict that same property for other structures not yet measured or even not yet prepared.<sup>3</sup>

Quantitative structure–retention relationships represent statistical models that quantify the relation between the structure of molecules and their chromatographic retention indices, allowing the prediction of the retention indices of novel compounds. QSRR on the retention indices have been reported for different types of organic compounds.<sup>4–8</sup>

The application of these techniques usually requires variable selection for building well-fitted models. In this work, the genetic algorithm selection method was employed for the variable selection in the MLR method. Nowadays, GA is well-known as an interesting and widely used variable selection method.<sup>9–11</sup>

The aim of this work was to search for an efficient method to build an accurate quantitative relationship between the molecular structure and the retention indices of *A. atlantica* essential oils by GA–MLR.

## EXPERIMENTAL

### *Computer hardware and software*

A Pentium IV personal computer (CPU at 3.06 GHz) with the Windows XP operating system was used. The geometry optimization was performed with HyperChem (Version 7.0 Hypercube, Inc). For the calculation of the molecular descriptors, Dragon 2.1 software was used.<sup>12</sup> SPSS software (version 11.50, SPSS, Inc.) was employed for the simple MLR analysis. The GA–MLR regression and the other calculations were performed in Matlab (Version 7.0, Math works, Inc).

### *Data set*

The data set of the GC retention indices was taken from the values reported by Laouer *et al.*<sup>13</sup> The Kovats retention indices were obtained according to the following equation:

$$I = 100N + 100 \left[ \frac{\log x_i - \log x_z}{\log x_{(z+1)} - \log x_z} \right] \quad (1)$$

where  $x$  refers to the adjusted retention volume or time,  $z$  is the number of carbon atoms of the *n*-alkane eluting before and  $z+1$  is the number of carbon atoms of the *n*-alkane eluting after the peak of interest. These values were obtained after subtracting the number of the carbons in the main chain ties 100 from Kovats indices ( $KI$ ).

The molecules in the data set, including the essential oils from *Ammoides atlantica*, are shown in Table I. The retention indices of all compounds were determined by GC and GC–MS under a single set of conditions. An apolar fused silica capillary column Perkin Elmer Elite-5 MS (5 % phenylmethylsiloxane, 30 m×0.25 mm, 0.25 µm film thickness) was used. The column oven temperature was programmed from 60–185 °C at 1.5 °C min<sup>-1</sup>, held isothermal at 185 °C for 1 min and then raised to 275 °C at 9 °C min<sup>-1</sup>.

The data set was split into a training set and a prediction set. The prediction set of 6 compounds was selected randomly from the original 32 of essential oil components, with the remaining compounds constituting the training set. The training set of 26 compounds, with *RI* values in the range 931–1582, was used to adjust the parameters of the model, and the test set of 6 compounds, with *RI* in the range 954–1545, was used to evaluate its predictive ability.

### *Determination of molecular descriptors*

Molecular descriptors are defined as numerical characteristics associated with chemical structures. The molecular descriptor is the final result of a logic and mathematical procedure which transforms chemical information encoded within a symbolic representation of a molecule into a useful number applied to correlate physical properties.

The Dragon software was used to calculate the descriptors in this research and a total of 1481 molecular descriptors, from 18 different types of theoretical descriptors, were calculated for each molecule. Since the values of many descriptors are related to the bonds length and bonds angles *etc.*, the chemical structure of every molecule must be optimized before calculating its molecular descriptors. For this reason, the chemical structure of the 32 studied molecules were drawn with Hyperchem software and saved with the HIN extension. To optimize the geometry of these molecules, the AM1 geometrical optimization was applied. After optimizing the chemical structures of all compounds, the molecular descriptors were calculated using Dragon. A wide variety of descriptors have been reported in the literature, having been used in QSAR analysis.<sup>14–19</sup>

### *Genetic algorithm for descriptor selection*

In QSPR studies, after calculating the molecular descriptors from optimized chemical structures of all the components available in the data set, the problem is to find a linear equation that can predict the desired property with the least number of variables as well as highest accuracy.

In other words, the problem is to find a subset of variables (most statistically effective molecular descriptors for the *RI*) from all the available variables (all molecular descriptors) that can predict *RI* with the minimum error in comparison to the experimental data.

A generally accepted method for this problem is the genetic algorithm based multivariate linear regression (GA-MLR). In this method, the genetic algorithm is applied for the selection of the best subset of variables with respect to an objective function. This algorithm was presented by Leardi *et al.* for the first time.<sup>20</sup>

The GA is a stochastic method to solve the optimization problems defined by fitness criteria, applying the evolution hypothesis of Darwin and different genetic functions, *i.e.*, crossover and mutation.

To select the most relevant descriptors, the evolution of the population was simulated.<sup>21–23</sup> The population of the first generation was selected randomly. Each individual member in the population, defined by a chromosome of binary values, represented a subset of descriptors. The number of the genes at each chromosome was equal to the number of the descriptors. A gene was given the value of 1, if its corresponding descriptor was included in the subset; otherwise, it was given the value of zero. The number of the genes with the value of 1 was kept relatively low to have a small subset of descriptors.<sup>24</sup> As a result, the probability of generating 0 for a gene was set greater (at least 60 %) than the value of 1. The operators used here were crossover and mutation. The application probability of these operators was varied linearly with a generation renewal (0.0–0.1 % for mutation and 60–90 % for crossover). The population size was varied between 50 and 250 for different GA runs. For a typical run, the evolution of the generation was stopped when 90 % of the generations took the same fitness.

### *MLR Modeling*

The general purpose of multiple regressions is to quantify the relationship between several independent or predictor variables and a dependent variable. A set of coefficients defines the single linear combination of independent variables (molecular descriptors) that best describes retention indices. The retention indices value for each essential oil component would then be calculated as a composite of each molecular descriptor weighted by the respective coefficients. A multi-linear model can be represented as:

$$y = \beta_0 + \beta_1 \chi_1 + \beta_2 \chi_2 + \beta_3 \chi_3 + \cdots + \beta_k \chi_k + \varepsilon \quad (2)$$

where  $k$  is the number of independent variables,  $\beta_1, \dots, \beta_k$  the regression coefficients and  $y$  is the dependent variable. The regression coefficients represent the independent contributions of each calculated molecular descriptor. The algebraic MLR model is defined in Eq. (1) and in matrix notation:

$$y = Xb + e \quad (3)$$

when  $X$  is of full rank, the least-squares solution is  $b = (X^T X)^{-1} X^T y$ , where  $b$  is the estimator for the regression coefficients in  $b$  and  $X^T$  is a transpose of  $X$ .

A single MLR model was developed for essential oil compounds using SPSS software (version 11.50, SPSS Inc., USA). The MLR model was built using a training set and validation using an external prediction set. Multiple linear regression (MLR) techniques based on least-

squares procedures are very often employed for estimating the coefficients involved in a model equation.<sup>25</sup>

## RESULTS AND DISCUSSION

### *MLR Analysis*

The multiple linear regression method (MLR) is one of the most used modeling methods in QSRR. Thus, MLR analysis was performed to derive the best QSAR model. A small number of molecular descriptors proposed by our team were used to establish a QSRR model. The MLR technique was performed on the molecules of the training set shown in Table I. After regression analysis, a few suitable models were obtained, from which the best model was selected and is presented in Eq. (3). A small number of molecular descriptors (*RDF035m*, *PCD* and *Q2*) proposed were used to establish the QSRR model. Additional validation was performed on an external data set consisting of 6 essential oil compounds. MLR Analysis provided a useful equation that can be used to predict the RI of essential oil based upon these parameters. The best equation obtained for the RI of the essential oil compounds is:

$$RI = 590.81 + 20.56RDF035m + 107.41PCD + 839.37Q2 \quad (4)$$

TABLE I. The data set and the corresponding observed and predicted *RI* values by GA-MLR for the training and test set (*E* – relative error)

No.	Compound	<i>RI</i> (Exp)	<i>RI</i> (GA-MLR)	<i>E</i> / %
Training set				
1	$\alpha$ -Thujene	931	993.31	6.69
2	$\alpha$ -Pinene	941	967.82	2.85
3	Sabinene	975	976.75	0.18
4	$\beta$ -Pinene	980	968.41	-1.18
5	$\alpha$ -Phellandrene	1007	1051.05	4.37
6	$\delta$ -3-Carene	1013	998.54	-3.38
7	<i>p</i> -Cymene	1026	1112.58	8.43
8	Limonene	1030	995.12	-3.38
9	$\gamma$ -Terpinene	1064	1010.16	-5.05
10	<i>cis</i> -Sabinene hydrate	1069	989.20	-7.46
11	Linalool	1099	1178.61	7.24
12	<i>cis-p</i> -Menth-2-en-1-ol	1123	1193.25	6.25
13	$\alpha$ -Terpineol	1186	1118.92	-5.65
14	Methyl thymol	1236	1290.56	4.41
15	Thymol	1292	1265.62	-2.04
16	Carvacrol	1297	1257.97	-3.00
17	$\beta$ -Caryophyllene	1420	1394.21	-1.81
18	$\alpha$ -Amorphene	1486	1490.01	0.26
19	$\gamma$ -Cadinene	1516	1399.89	-7.65
20	$\delta$ -Cadinene	1527	1517.06	-0.65
21	Germacrene B	1560	1572.98	0.83
22	Caryophyllene oxide	1582	1617.89	2.26



TABLE I. Continued

No.	Compound	<i>RI</i> (Exp)	<i>RI</i> (GA-MLR)	<i>E</i> / %
		Prediction set		
1	Camphene	954	1039.2	8.93
2	Myrcene	993	997.8	0.48
3	$\alpha$ -Terpinene	1020	1012.2	-0.76
4	1,8-Cineole	1032	1082.3	4.88
5	Terpinolene	1088	985	-9.45
6	Terpinen-4-ol	1177	1119.5	-4.87
7	Methyl carvacrol	1244	1272.5	2.29
8	$\alpha$ -Copaene	1379	1382.8	0.28
9	$\alpha$ -Murolene	1501	1482.9	-1.19
10	Selina-3,7(11)-diene	1545	1406.2	-8.98

Positive values of the regression coefficients indicate that the indicated descriptor contributes positively to the value of *RI*, whereas negative values indicate that the greater the value of the descriptor, the lower is the value of *RI*. In other words, increasing *RDF035m*, *PCD* and *Q2* will increase the extent of the *RI* of essential oil compounds.

For an evaluation of the predictive power of the generated MLR, the optimized model was applied for prediction of the *RI* values of 6 compounds in the prediction set, which were not used in the optimization procedure. For the constructed models, the predictive ability of the MLR model was evaluated by calculation of statistical parameters.

The data set and the corresponding experimental and predicted *RI* values of all the molecules studied in this work are summarized in Table I. Plots of the values predicted by the GA-MLR against the experimental values of the retention indices of the training and prediction sets are shown in Fig. 1. The residuals (experimental *RI* – predicted *RI*) obtained by the GA-MLR modeling *versus* the experimental *RI* values are shown in Fig. 2. The distribution of the residuals on both sides of the zero line indicates there is no systematic error in the GA-MLR model.

#### Statistical parameters

For an evaluation of the predictive power of the generated MLR, the optimized model was applied for the prediction of the *RI* values of the test compounds in the prediction set, which were not used in the optimization procedure. For the constructed models, two general statistical parameters were selected to evaluate the prediction ability of the model for *RI* values. For this case, the predicted *RI* of each sample in the prediction step was compared with the experimental *RI*.

The root mean square error of prediction (*RMSEP*) is a measurement of the average difference between predicted and experimental values, at the prediction

stage. The *RMSEP* can be interpreted as the average prediction error, expressed in the same units as the original response values. The *RMSEP* was obtained using the following formula:

$$RMSEP = \frac{100}{\bar{y}} \left[ \frac{1}{n} \sum_{i=1}^n (y_i - \hat{y}_i)^2 \right]^{0.5} \quad (5)$$

The second statistical parameter was the relative error of prediction (*REP*) that shows the predictive ability of each component, and is calculated as:

$$REP (\%) = \frac{100}{\bar{y}} \left[ \frac{1}{n} \sum_{i=1}^n (\hat{y}_i - y_i)^2 \right]^{0.5} \quad (6)$$

where  $y_i$  is the experimental *RI* value of the essential oil in the sample  $i$ ,  $\hat{y}_i$  represents the predicted *RI* value of the essential oil in the sample  $i$ ,  $\bar{y}$  is the mean of experimental *RI* values in the prediction set and  $n$  is the total number of samples used in the prediction set.

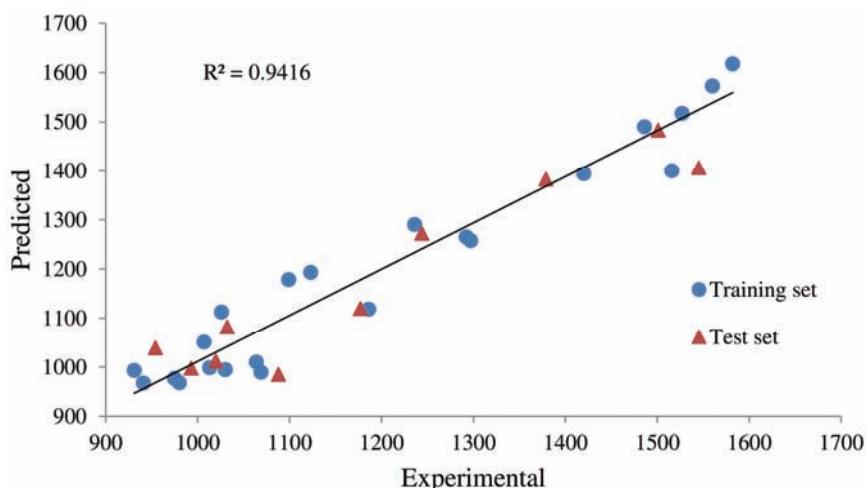


Fig. 1. The *RI* values predicted by the MLR modeling vs. the experimental *RI* values.

The statistical parameters calculated for the GA-MLR model are listed in Table II.

Topliss and Edwards<sup>26</sup> demonstrated that the more independent variables screened, the more independent variables are available for selection in a multiple linear regression model and the more likely a model will be found by chance. These authors recommended that in order to reduce the risk of chance correlations, there should be a certain ratio of data points to the number of independent variables available. A solution for the detection of a chance correlation is the *Y*-randomization test.<sup>27</sup> In order to assess the robustness of the model, the *Y*-ran-

domization test was applied in this study. The dependent variable vector ( $RI$ ) was randomly shuffled and a new QSPR model was developed using the original independent variable matrix. The new QSPR models (after several repetitions) are expected to have low  $R^2$  and  $Q^2$  values (Table III). If the  $R^2$  and  $Q^2$  values of these models were much lower than those of the original model, it could be considered that the model was reasonable and had not been obtained by the chance.

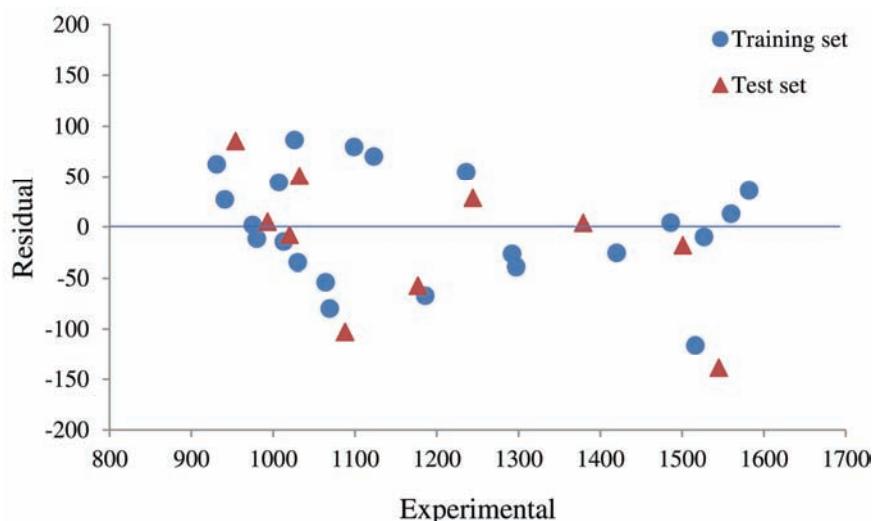


Fig. 2. Plot of the residuals against the experimental values of the retention indices.

TABLE II. Statistical parameters obtained by application of the MLR method to the test set

Parameter	Value
$RMSEP$	66.473
$REP / \%$	5.570
$R^2$	0.894
Number of descriptors	3

TABLE III.  $R^2$  train and  $Q^2$  LOO values after several  $Y$ -randomization tests

Iteration	$R^2$	$Q^2$
1	0.1568	0.0031
2	0.1217	0.0013
3	0.2186	0.0122
4	0.0729	0.0096
5	0.1354	0.0197
6	0.0097	0.2751
7	0.1877	0.0026
8	0.3235	0.1193
9	0.0814	0.0506
10	0.2099	0.0059

### Description of the models descriptors

The retention indices in GC depend on the relative solubility of the solute in the mobile and stationary phases, which depend on the molecular structure and chemical properties of the solute. Differences between these properties govern the retention behavior through the column.

In this work, three descriptors were selected including *RDF035m*, *PCD* and *Q2* for the prediction of the retention indices values. A brief description of the descriptors is presented in Table IV. The methods for the calculations of these descriptors and their meaning have been explained in the Handbook of Molecular Descriptors by Todeschini *et al.*<sup>14</sup>

TABLE IV. A brief description of the descriptors used in this study

No.	Descriptor	Definition
1	<i>RDF035m</i>	Radial distribution function – 3.5/weighted by atomic mass
2	<i>PCD</i>	Difference of multiple path counts to path counts
3	<i>Q2</i>	Total squared charge

The first descriptor is *RDF035m*, which is one of the radial distribution function (*RDF*) descriptors. A *RDF* in this form meets all the requirements for a 3D structure descriptor. It is independent of the atom number (*i.e.*, the size of a molecule), it is unique regarding the three-dimensional arrangement of the atoms and it is invariant against translation and rotation of the entire molecule.

Formally, a radial distribution function of an ensemble of  $N$  atoms can be interpreted as the probability distribution to find an atom in a spherical volume of radius  $r$ .<sup>21</sup> The following equation represents the radial distribution function code as it was used in this investigation:

$$g(r) = f \sum_i^{N-1} \sum_j^{N} A_i A_j e^{-B(r-r^4)^2} \quad (7)$$

where  $f$  is a scaling factor and  $N$  is the number of atoms. By including characteristic atomic properties  $A$  of the atoms  $i$  and  $j$ , the *RDF* codes can be used in different tasks to fit the requirements of the information to be represented. The exponential term contains the distance  $r_{ij}$  between the atoms  $i$  and  $j$  and a smoothing parameter  $B$ , which defines the probability distribution of the individual distances.  $g(r)$  was calculated at a number of discrete points with defined intervals. The atomic properties  $A_i$  and  $A_j$  used in this equation enable the discrimination of the atoms of a molecule for almost any property that can be attributed to an atom. Such a distribution function provides, besides information about interatomic distances in the whole molecule, the opportunity to gain access to other valuable information, *e.g.*, bond distance, ring types, planar and non-planar systems and atom types. This fact is a most valuable consideration for a computer-assisted

code elucidation. The radial distribution function in this form meets the entire requirement mentioned above, especially the invariance against linear translations.<sup>28</sup>

Additionally, the *RDF* descriptors can be restricted to specific atom types or distance ranges to represent specific information in a certain three-dimensional structure space (*e.g.*, to describe the steric hindrance or the structure/activity properties of a molecule). *RDF035m* displays a positive sign, which indicates that a retention index is directly related to this descriptor.

The second descriptor of this model was the difference of multiple path counts to path counts (*PCD*) that describes information related to bonds and distances (bond orders, saturation and ratio of multiple bonds to single bonds).<sup>14</sup> Its effect on the retention indices was positive, which indicates that the retention time is directly related to this descriptor.

The final descriptor of this model was the total squared charge (*Q2*)<sup>29–31</sup> that is obtained by the following formula:

$$Q2 = \sum_i^N Q_i^2 \quad (8)$$

where  $Q_i$  is the amount of charge on atom  $i$  and  $N$  is the number of atoms of that molecule. This descriptor had a positive effect on the retention indices.

The correlation coefficient matrix for the descriptors used in this study (GA-MLR), is listed in Table V. The data indicate there is a very low correlation between the descriptors used in this research. Therefore, the ability of the resulting QSRR regression models to enable accurate prediction of the retention indice is not related to co-linearity between the variables.

TABLE V. The correlation coefficient matrix for the descriptors used in this study

	<i>PCD</i>	<i>Q2</i>	<i>RDF035m</i>
<i>PCD</i>	1	–	–
<i>Q2</i>	0.259	1	–
<i>RDF035m</i>	0.108	0.532	1

#### CONCLUSIONS

In this paper, a simple QSRR model was presented for the prediction of the *RI* of essential oils. This model is a multivariate linear model, which has three variables (molecular descriptors). These three molecular descriptors were selected using the GA-MLR technique. These variables are calculated based on the chemical structure of the molecules. The QSRR model with the simply calculated molecular descriptors could be employed to estimate the retention indices for new compounds, even in the absence of standard candidates.

## ИЗВОД

КВАНТИТАТИВНА РЕЛАЦИЈА ИЗМЕЂУ СТРУКТУРЕ И РЕТЕНЦИЈЕ ЗА  
ПРЕДВИЋАЊЕ ИНДЕКСА РЕТЕНЦИЈЕ ЕСЕНЦИЈАЛНИХ УЉА  
ИЗ *Ammoides atlantica*

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Развијен је једноставан, дескриптиван и интерпретативан модел, заснован на квантитативној релацији између структуре и ретенције (QSRR), уз примену генетичког алгоритма са линеарном регресијом (A-MLR). Помоћу тог модела предвиђају се ретенциони индекси (*RI*) за компоненте есенцијалних уља. Помоћу молекулског моделовања идентификована су три битна дескриптора за опис *RI*. Састављена је база података која се састоји из *RI* за 32 молекула који су садржани у есенцијалним уљима, од више од 931 познатих. Затим је израчунат погодан скуп молекулских дескриптора и најбитнији међу њима су одређени помоћу GA-MLR. Добијен је модел са ниском грешком и добрим кофицијентом корелације. Модел је онда употребљен за предвиђање *RI* вредности за неке компоненте есенцијалних уља које нису коришћене за добивање QSRR модела.

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## REFERENCES

1. N. Achak, A. Romane, M. Alifriqui, R. P. Adams, *J. Essent. Oil Res.* **20** (2008) 200
2. H. Laouer, M. M. Zerroug F. Sahli, A. N. Chaker, G. Valentini, G. Ferretti, M. Grande, J. Anaya, *J. Essent. Oil Res.* **15** (2003) 135
3. A. R. Katritzky, D. G. Fara, *Energy Fuels* **19** (2005) 922
4. M. Jalali-Heravi, M. H. Fatemi, *J. Chromatogr. A* **915** (2001) 177
5. Z. Garakani-Nejad, M. Karlovits, W. Demuth, T. Stimpfl, W. Vycudilik, M. Jalali-Heravi, K. Varmuza, *J. Chromatogr. A* **1028** (2004) 287
6. J. Acevedo-Martinez, J. C. Escalona-Arranz, A. Villar-Rojas, F. Tellez-Palmero, P. R. Roses, L. Gonzalez, R. Carrasco-Velar, *J. Chromatogr. A* **1102** (2006) 238
7. P. Tulasamma, K. S. Reddy, *J. Mol. Graphics Modell.* **25** (2006) 507
8. K. Heberger, T. Kowalska, *Chemom. Intell. Lab. Syst.* **47** (1999) 205
9. U. Depczynski, V. J. Frost, K. Molt, *Anal. Chim. Acta* **420** (2000) 217
10. B. K. Alsberg, N. Marchand-Geneste, R. D. King, *Chemom. Intell. Lab. Syst.* **54** (2000) 75
11. D. Jouanrimbaud, D. L. Massart, R. Leardi, O. E. De Noord, *Anal. Chem.* **67** (1995) 4295
12. Dragon 6, R. Todeschini, V. Consonni, M. Pavana, available at [http://www.talete.mi.it/products/dragon\\_description.htm](http://www.talete.mi.it/products/dragon_description.htm)
13. H. Laouer, N. Boulaacheb, S. Akkal, G. Singh, P. Marimuthu, C. De Heluani, C. Catalan, N. Baldovini, *J. Essent. Oil Res.* **20** (2008) 266
14. R. Todeschini, V. Consonni, *Handbook of Molecular Descriptors*, Wiley-VCH, Weinheim, Germany, 2000.
15. L. B. Kier, L. H. Hall, *Molecular Connectivity in Structure–Activity Analysis*, RSP-Wiley, Chichester, UK, 1986.
16. E. V. Kostantinova, *J. Chem. Inf. Comp. Sci.* **36** (1997) 54



17. G. Rucker, C. Rucker, *J. Chem. Inf. Comp. Sci.* **33** (1993) 683
18. J. Galvez, R. Garcia, M. T. Salabert, R. Soler, *J. Chem. Inf. Comp. Sci.* **34** (1994) 520
19. P. Broto, G. Moreau, C. Vandicke, *J. Med. Chem.* **19** (1984) 66
20. R. Leardi, R. Boggia, M. Terrile, *J. Chemom.* **6** (1992) 267
21. J. Hunger, G. Huttner, *J. Comput. Chem.* **20** (1999) 455
22. S. Ahmad, M. M. Gromiha, *J. Comput. Chem.* **24** (2003) 1313
23. C. L. Waller, M. P. Bradley, *J. Chem. Inf. Comput. Sci.* **39** (1999) 345
24. J. Aires-de-Sousa, M. C. Hemmer, J. Casteiger, *Anal. Chem.* **74** (2002) 80
25. J. N. Miller, J. C. Miller, *Statistics and Chemometrics for Analytical Chemistry*, Prentice Hall, London, 2000
26. J. G. Topliss, R. P. Edwards, *J. Med. Chem.* **22** (1979) 1238
27. A. Tropsha, P. Gramatica, V. K. Gombar, *QSAR Comb. Sci.* **22** (2003) 69
28. M. P. Gonzalez, Z. Gandara, Y. G. Gomez, *Eur. J. Med. Chem.* **43** (2008) 1360
29. N. Bodor, Z. Gabanyi, C. K. Wong, *J. Am. Chem. Soc.* **111** (1989) 3783
30. L. Buydens, D. Massart, P. Geerlings, *Anal. Chem.* **55** (1983) 738
31. M. Karelson, S. Victor, *Chem. Rev.* **96** (1996) 1027.